RESEARCH

Open Access

Risk factors and mortality in children with severe pertussis: the role of exchange transfusion in a PICU



Junming Huo¹, Song Chen¹, Yanran Qin¹, Feng Xu¹ and Chenjun Liu^{1*}

Abstract

Objective Although multiple risk factors have been reported for adverse outcomes in children with severe pertussis, their predictive values and the benefits of interventions such as exchange transfusion remain poorly understood. Therefore, we aimed to comprehensively evaluate the risk factors associated with mortality in children with severe pertussis and assess the potential benefits of exchange transfusion therapy.

Methods A retrospective analysis of 170 pertussis patients admitted to the Pediatric Intensive Care Unit (PICU) between January 2018 and June 2024 was performed.

Results Among the 170 patients, 38 (22.35%) died. The death group exhibited significantly higher white blood cell (WBC) counts (67.31 vs. 28.41 × 10^9/L, P < 0.001), neutrophils (29.95 vs. 11.61 × 10^9/L, P < 0.001), and C-reactive protein (CRP) (29 vs. 8 mg/L, P < 0.001). Additionally, sepsis (39.47% vs. 9.09%, P < 0.001), shock (63.16% vs. 6.06%, P < 0.001), ARDS (23.68% vs. 2.27%, P < 0.001), and acute kidney injury (21.05% vs. 0.76%, P < 0.001) were more prevalent in the death group. ROC analysis showed that WBC counts had a predictive value for mortality (AUC = 0.75, sensitivity = 0.78, specificity = 0.68), with an optimal cutoff of 48.58 × 10^9/L.

Conclusion High WBC counts are significantly correlated with increased mortality risk in severe pertussis children, with a threshold of 48.58 × 10^9/L marking high risk. Although exchange transfusion can reduce WBC counts and improve symptoms, its benefit is limited in patients with severe secondary infections, necessitating tailored treatment strategies.

Keywords Pertussis, Risk factors, Exchange transfusion, Children, Mortality, PICU

Introduction

Pertussis is an acute infectious respiratory disease caused by *Bordetella pertussis* and is one of the top ten leading causes of death in children under the age of one worldwide. The infection rate is highest in infants, particularly those under 6 months of age, with the majority of deaths occurring in infants under 3 months. It is estimated that up to 400,000 pertussis-related deaths occur annually, with 90% of these deaths happening in developing countries [1–3]. Severe pertussis is a common cause of hospitalization in infants, accompanied by a high risk of mortality, placing a significant burden on healthcare systems [4, 5]. Many studies [6–8] have indicated that high white blood cell (WBC) counts are a risk factor for death in patients with pertussis, and some case reports [9–11] and studies [12, 13] begun to explore interventions such as exchange transfusions to address this issue.



© The Author(s) 2025. **Open Access** This article is licensed under a Creative Commons Attribution-NonCommercial-NoDerivatives 4.0 International License, which permits any non-commercial use, sharing, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons licence, and indicate if you modified the licensed material. You do not have permission under this licence to share adapted material derived from this article or parts of it. The images or other third party material in this article are included in the article's Creative Commons licence, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons licence and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this licence, visit http://creativecommons.org/licenses/by-nc-nd/4.0/.

^{*}Correspondence:

Chenjun Liu

liucwd@163.com

¹ Department of Critical Care Medicine, National Clinical Research Center for Child Health and Disorders, Ministry of Education Key Laboratory of Child Development and Disorders, Chongqing Key Laboratory of Pediatrics Metabolism and Inflammatorydiseases, Children'S Hospital of Chongqing Medical University, 136 Zhongshan 2 Road, Yuzhong District, Chongqing 400014, China

However, case reports and current research sample sizes— with the largest being 23 [14]—still lack strong persuasive power, making it difficult to draw consistent conclusions. Furthermore, discrepancies exist among studies regarding the specific WBC counts thresholds for predicting death and survival, indicating that our understanding of this important biomarker remains insufficient.

To further investigate the risk factors for mortality in severe pertussis, establish the optimal predictive value of high WBC counts in this condition, and evaluate the potential for improving prognosis through exchange transfusion therapy, we plan to conduct a larger case– control study with a more systematic design, aimed at validating and exploring effective intervention strategies.

Methods

Study design

Retrospective analysis of clinical data and prognostic outcomes of pertussis patients admitted to the Pediatric Intensive Care Unit (PICU) of Chongqing Medical University Children's Hospital from January 2018 to June 2024. The Institutional Review Committee of Chongqing Medical University Children's Hospital has approved this study. The informed consent was waived due to the retrospective design.

The diagnosis of pertussis was based on clinical manifestations and laboratory test results (positive PCR for *Bordetella pertussis*). Patients were identified by retrieving discharge diagnosis data from electronic medical databases. The inclusion criteria include patients admitted to the pediatric intensive care unit (PICU) due to severe pertussis. Excluded patients with severe underlying diseases (leukemia, liver failure, etc.) and those who originally had severe infections and pertussis infection was not the initial infection.

Data quality control and management

The data for this study were retrieved from the hospital's data center and collected using Excel spreadsheets. A dedicated personnel member managed the data, which were stored on a secure computer. Strict quality control measures were implemented to ensure data accuracy and completeness.

Data collection and definition of variables

Demographic and clinical data were collected, including age, gender, diagnosis(AKI,Acute kidney injury; ARDS, Acute respiratory distress syndrome;PH,pulmonary arterial hypertension; epilepsy, hypoproteinemia;sepsis; shock; respiratory failure; myocardial injury; hepatic injury) as well as the highest white blood cell count (WBC) before treatment (including lymphocyte absolute value, monocyte absolute value, neutrophil absolute value), platelet count (PLT), C-reactive protein (CRP), procalcitonin (PCT), alanine transaminase (ALT), bilirubin, creatinine, and laboratory tests for other respiratory pathogen infections. Laboratory specimens are taken upon admission. The WBC from the last test taken during hospitalization were also collected.

Imaging results were categorized into pneumonia, atelectasis, and lung consolidation. The treatment needs of patients during hospitalization were divided into use of vasoactive drugs, albumin, hormones, intravenous immune globulin (IVIG), extracorporeal membrane oxygenation (ECMO), ventilator, high frequency ventilation, invasive ventilation, exchange Transfusion (Double Blood Volume, a procedure where the patient's blood volume is replaced with donor blood). The primary focus of this study was to evaluate the frequency of mortality at any point.

Statistical analysis

This study used R 4.4.1 statistical analysis software, where the continuous data with skewed distribution was represented by M (IQR), and the rank sum test of two independent samples was used; The classified data is represented by n (%) and Chi-squared or Fisher's exact probability method is used. The difference between groups is considered statistically significant with P < 0.05. Patients are divided into Survivors group or Death group based on outcomes and Without or With groups based on whether they had exchange transfusion.

Results

A total of 170 children were included in this study (Table 1), with 132 (77.65%) survivors and 38 (22.35%) deaths.

Compared to survivors, the Death group had significantly higher WBC counts (67.31 vs. $28.41 \times 10^{\circ}/L$, P < 0.001), lymphocytes (24.47 vs. $14.62 \times 10^{\circ}/L$, P = 0.034), neutrophils (29.95 vs. $11.61 \times 10^{\circ}/L$, P < 0.001), monocytes (2.90 vs. $1.23 \times 10^{\circ}/L$, P < 0.001), and CRP levels (29 vs. 8 mg/L, P < 0.001). The PCT (0.22 vs. 0.11 ng/mL, P < 0.001) and bilirubin (9.30 vs. 6.00μ mol/L, P = 0.035) were also higher in the Death group, while PLT was lower (524 vs. $584 \times 10^{\circ}/L$, P = 0.021).

The incidence of sepsis (39.47% vs. 9.09%, P < 0.001), shock (63.16% vs. 6.06%, P < 0.001), ARDS (23.68% vs. 2.27%, P < 0.001), and AKI (21.05% vs. 0.76%, P < 0.001) was significantly higher in the Death group. Regarding treatment, the Death group had a higher proportion of vasoactive drug use (92.11% vs. 47.73%, P < 0.001), albumin administration (92.11% vs. 67.42%, P < 0.001), ECMO support (7.89% vs. 0%, P = 0.010), and exchange transfusion (42.11% vs. 17.42%, P = 0.001). However, IVIG use

Table 1 Characteristics of patients according to outcome

Variables	Total (<i>n</i> = 170)	Survivors (n = 132)	Death (n = 38)	Р	
Age (day), M (IQR)	84 (49, 151)	84 (50, 161)	83 (48, 146)	0.599	
Gender (male), n(%)	90 (52.94)	74 (56.06)	16 (42.11)	0.129	
Immunization, n(%)	76 (44.71)	59 (44.70)	17 (44.74)	0.997	
WBC (× 10 ⁹ /I),M (IQR)	32.77 (17.68, 58.25)	28.41 (16.01, 47.36)	67.31 (29.31, 92.76)	<.001	
Lymphocytic absolute value, M (IQR)	15.18 (7.68, 27.20)	14.62 (7.22, 24.04)	24.47 (9.79, 32.10)	0.034	
Neutrophils absolute value, M (IQR)	13.62 (7.20, 23.83)	11.61 (6.37, 19.13)	29.95 (14.52, 49.94)	<.001	
Monocytes absolute value, M (IQR)	1.44 (0.73, 2.80)	1.23 (0.70, 2.15)	2.90 (1.25, 4.87)	<.001	
PLT (× 10 ⁹ /l), M (IQR)	565 (466, 708)	584 (481, 715)	524 (330, 666)	0.021	
CRP (mg/l), M (IQR)	8 (8, 22)	8 (8, 14)	29 (8, 45)	<.001	
PCT (ng/ml), M (IQR)	0.11 (0.07, 0.31)	0.10 (0.07, 0.18)	0.22 (0.11, 0.63)	<.001	
Bilirubin (μmol/l), M (IQR)	6.50 (2.00, 13.68)	6.00 (1.90, 12.07)	9.30 (4.20, 21.92)	0.035	
Creatinine (µmol/l), M (IQR)	21.50 (17.52, 25.48)	21.00 (17.90, 25.20)	22.20 (16.00, 26.30)	0.830	
ALT (U/L), M (IQR)	34.00 (23.85, 47.00)	33.15 (23.60, 46.00)	37.70 (26.10, 53.85)	0.205	
PCO ₂ (mmHg), M (IQR)	43.30 (37.00, 52.00)	43.00 (37.00, 49.60)	50.75 (40.25, 56.90)	0.050	
PO ₂ (mmHg), M (IQR)	86.40 (65.03, 118.50)	86.60 (65.07, 111.25)	85.00 (65.25, 148.00)	0.440	
PH, n(%)	18 (10.59)	16 (12.12)	2 (5.26)	0.362	
Epilepsy, n(%)	8 (4.71)	7 (5.30)	1 (2.63)	0.802	
Hypoproteinemia, n(%)	74 (43.53)	58 (43.94)	16 (42.11)	0.841	
Sepsis, n(%)	27 (15.88)	12 (9.09)	15 (39.47)	<.001	
Shock, n(%)	32 (18.82)	8 (6.06)	24 (63.16)	<.001	
Respiratory failure, n(%)	164 (96.47)	128 (96.97)	36 (94.74)	0.874	
ARDS, n(%)	12 (7.06)	3 (2.27)	9 (23.68)	<.001	
Myocardial injury, n(%)	50 (29.41)	35 (26.52)	15 (39.47)	0.122	
Hepatic injury, n(%)	34 (20.00)	24 (18.18)	10 (26.32)	0.269	
AKI, n(%)	9 (5.29)	1 (0.76)	8 (21.05)	<.001	
Other respiratory pathogen infections ^a , n(%)	129 (75.88)	97 (73.48)	32 (84.21)	0.173	
Pneumonia, n(%)	165 (97.06)	127 (96.21)	38 (100.00)	0.501	
Atelectasis, n(%)	66 (38.82)	54 (40.91)	12 (31.58)	0.298	
Lung consolidation, n(%)	121 (71.18)	92 (69.70)	29 (76.32)	0.427	
Vasoactive drugs, n(%)	98 (57.65)	63 (47.73)	35 (92.11)	<.001	
Albumin, n(%)	124 (72.94)	89 (67.42)	35 (92.11)	0.003	
Hormones, n(%)	98 (57.65)	78 (59.09)	20 (52.63)	0.478	
IVIG, n(%)	110 (64.71)	91 (68.94)	19 (50.00)	0.031	
ECMO, n(%)	3 (1.76)	0 (0.00)	3 (7.89)	0.010	
Ventilator, n(%)	155 (91.18)	117 (88.64)	38 (100.00)	0.064	
High frequency ventilation, n(%)	11 (6.47)	7 (5.30)	4 (10.53)	0.436	
Invasive ventilation, n(%)	142 (83.53)	107 (81.06)	35 (92.11)	0.106	
Exchange transfusion, n(%)	39 (22.94)	23 (17.42)	16 (42.11)	0.001	

ALT Alanine transaminase, AKI Acute kidney injury, ARDS Acute respiratory distress syndrome, CRP C-reactive protein, ECMO Extracorporeal membrane oxygenation, IQR Interquartile range, IVIG Intravenous immune globulin, M Median, PCT Procalcitonin, PCO₂ Partial pressure of carbon dioxide, PO₂ Oxygen partial pressure, PLT Platelet, PH Pulmonary arterial hypertension, WBC White blood cells

^a Other respiratory pathogen infections include the following pathogens: Adenovirus, Epstein-Barr Virus, Bocavirus, Parainfluenza Virus, Coronavirus, Respiratory Syncytial Virus, Influenza Virus, Acinetobacter baumannii, Citrobacter freundii, Enterobacter cloacae, Escherichia coli, Klebsiella pneumoniae, Streptococcus pneumoniae, Haemophilus influenzae, Staphylococcus aureus, Moraxella catarrhalis, Enterobacter aerogenes, Serratia marcescens, Leminospira nodosa, Methicillin-Resistant Staphylococcus aureus, Elizabethkingia meningoseptica, Klebsiella oxytoca, Achromobacter xylosoxidans, Pseudomonas aeruginosa, Lysinibacillus sphaericus, Candida albicans, Chlamydia pneumoniae, Mycoplasma pneumoniae, Chlamydia trachomatis, and others

was lower in the Death group (50.00% vs. 68.94%, P = 0.031).

Patients who underwent exchange transfusion had higher WBC counts (66.12 vs. 25.42×10^9 /L, *P* < 0.001), lymphocytes (28.22 vs. 12.11×10^9 /L, *P* < 0.001), neutrophils (27.57 vs. 10.26×10^9 /L, *P* < 0.001), monocytes (2.84 vs. 1.09×10^9 /L, *P* < 0.001), and CRP (18 vs. 8 mg/L, *P* = 0.001), as well as a higher incidence of shock (48.72% vs. 9.92%, *P* < 0.001) and increased mortality (41.03% vs. 16.79%, *P* = 0.001) compared to those without exchange transfusion (Table 2).

Among patients who received exchange transfusion, the Death group had significantly higher WBC counts (80.06 vs. 62.40 ×10⁹/L, P= 0.017), neutrophils (42.75 vs. 22.47 ×10⁹/L, P< 0.001), monocytes (2.90 vs. 1.23 ×10⁹/L, P< 0.001), and CRP (32 vs. 8 mg/L, P= 0.019), whereas lymphocyte levels showed no significant difference (P = 0.944). And there also had a higher proportion of sepsis patients (31.25% vs. 4.35%, P= 0.033), shock patients (48.72% vs. 9.92%, P< 0.001) and more vasoactive drugs (74.36% vs. 52.67%, P= 0.016) use. In addition, the PO2 of the Death group (87.05 vs. 65.10 mmHg, P= 0.009) was higher than that of the Survivors group (Table 3).

In the ROC curve analysis, WBC counts demonstrated a predictive value for mortality, with an AUC of 0.75 (95% CI: 0.64–0.85). The optimal cutoff value was 48.58 $\times 10^{9}$ /L, yielding an accuracy of 0.76, sensitivity of 0.78, and specificity of 0.68 (Fig. 1 Diagnostic Performance of WBC: ROC Curve Analysis for Mortality Prediction).

Univariate analysis identified sepsis (OR = 6.52, 95% CI: 2.70–15.73, P < 0.001) and shock (OR = 26.57, 95% CI: 10.05–70.26, P < 0.001) as significant predictors of mortality (Table 4). In the multivariate model, both remained significant, with sepsis (OR = 3.79, 95% CI: 1.21–11.91, P = 0.023) and shock (OR = 13.88, 95% CI: 4.72–40.82, P < 0.001). WBC counts also remained a significant predictor (OR = 1.02, 95% CI: 1.01–1.04, P < 0.001).

Table 5 displayed the highest WBC levels recorded prior to treatment and the results of the final test for all patients categorized into Survivors and Death groups who underwent exchange transfusion. Supplementary Table 1 provided clinical characteristics of patients who received ECMO treatment in comparison to those who did not.

Discussion

We conducted a retrospective analysis of children with severe pertussis admitted to the PICU over the past 6 years. Our results revealed that sepsis, shock, and elevated WBC counts significantly increased the risk of adverse outcomes. In addition, patients receiving exchange transfusion had more severe conditions with higher mortality; among these patients, the proportions of sepsis and shock—as well as elevated WBC counts were significantly greater in those who died WBC counts

were significantly greater in those who died. WBC counts also demonstrated good predictive ability for distinguishing between death and survival (AUC of 0.76, 95% CI: 0.69–0.82) with an optimal cutoff value of 48.58.

The present study identified a mortality rate of 22% (38/170). This mortality rate is somewhat higher compared to other reports. One study reported a mortality of 14% among 49 children with pertussis in intensive care, while a 2021 study noted 13 deaths among 38 ICU admissions [15, 16]. These differences may be attributable to variations in patient populations, clinical settings, or treatment protocols.

One of the central findings of our study is the association between elevated WBC counts and mortality. Consistent with previous reports, the WBC counts in the death group were significantly higher than those in survivors [17]. Specifically, WBC counts of $\geq 30 \times 10^{9}/L$ or $\geq 50 \times 10^{9}/L$ were significantly associated with mortality, and when the WBC count exceeded $70 \times 10^{9}/L$, the odds ratio for mortality soared to 230.66 [15, 18, 19]. In our study, ROC curve analysis identified an optimal cutoff value of $48.58 \times 10^{9}/L$ (AUC = 0.76, 95% CI: 0.69–0.82). Differences in thresholds among studies may be attributed to variations in population age (this study mainly focused on infants under 3 months) or differences in the timing of treatment.

Secondary infections also emerged as a critical factor in our cohort. We observed that patients in the death group not only had higher neutrophil counts and CRP levels, but also a greater incidence of sepsis compared to survivors. Similar findings have been reported by other researchers [20, 21]. Even after exchange transfusion which effectively reduced overall WBC counts—the persistence of elevated inflammatory markers suggests that secondary infections may continue to drive adverse outcomes. This is further supported by the high rate of respiratory infections (up to 75.88% in our cohort) and comparable findings in other studies [22].

Exchange transfusion therapy for severe pertussis patients is based on theoretical foundations. Some researchers suggest that an increase in white blood cell counts may impair pulmonary microcirculation, leading to pulmonary arterial hypertension, heart failure, and hypoxemia [23, 24]. However, Winter et al. pointed out that pertussis toxin might cause more extensive and destructive damage to cardiac and respiratory functions by inhibiting key Gi protein signaling pathways in the heart and lungs [6]. They proposed that leukocytosis may simply be a marker of severe pertussis toxin (PT)related disease rather than a direct cause of death, yet exchange transfusion could still be beneficial by reducing

Table 2 Characteristics of patients according to outcome

Variables	Total (<i>n</i> = 170)	Without (<i>n</i> = 131)	With (<i>n</i> = 39)	Р
Age (day), M (IQR)	84 (49, 151)	85 (48, 141)	71 (51, 192)	0.732
Gender (male), n(%)	90 (52.94)	73 (55.73)	17 (43.59)	0.183
Immunization, n(%)	76 (44.71)	59 (45.04)	17 (43.59)	0.873
WBC (× 10 ⁹ /l),M (IQR)	32.77 (17.68, 58.25)	25.42 (15.38, 42.33)	66.12 (55.26, 83.10)	<.001
Lymphocytic absolute value, M (IQR)	15.18 (7.68, 27.20)	12.11 (6.14, 21.66)	28.22 (23.40, 37.78)	<.001
Neutrophils absolute value, M (IQR)	13.62 (7.20, 23.83)	10.26 (6.29, 16.92)	27.57 (20.33, 39.46)	<.001
Monocytes absolute value, M (IQR)	1.44 (0.73, 2.80)	1.09 (0.66, 1.94)	2.84 (2.12, 4.57)	<.001
PLT (× 10 ⁹ /l), M (IQR)	565(466, 708)	548 (446, 690)	620 (527, 743)	0.064
CRP (mg/l), M (IQR)	8 (8, 22)	8 (8, 15)	18 (8, 34)	0.001
PCT (ng/ml), M (IQR)	0.11 (0.07, 0.31)	0.11 (0.07, 0.30)	0.13 (0.08, 0.33)	0.278
Bilirubin (µmol/l), M (IQR)	6.50 (2.00, 13.68)	7.00 (2.00, 13.55)	6.30 (3.00, 13.15)	0.956
Creatinine (µmol/l), M (IQR)	21.50 (17.52, 25.48)	22.00 (18.00, 25.75)	21.00 (16.00, 25.20)	0.162
ALT (U/L), M (IQR)	34.00 (23.85, 47.00)	35.60 (26.25, 48.15)	30.60 (19.50, 42.70)	0.029
PCO ₂ (mmHg), M (IQR)	43.30 (37.00, 52.00)	43.00 (36.50, 50.25)	49.00 (40.40, 55.05)	0.03
PO ₂ (mmHg), M (IQR)	86.40 (65.03, 118.50)	89.10 (67.35, 124.50)	78.20 (61.50, 97.20)	0.142
PH, n(%)	18 (10.59)	15 (11.45)	3 (7.69)	0.709
Epilepsy, n(%)	8 (4.71)	5 (3.82) 3 (7.69)		0.567
Hypoproteinemia, n(%)	74 (43.53)	58 (44.27)	16 (41.03)	0.719
Sepsis, n(%)	27 (15.88)	21 (16.03)	6 (15.38)	0.923
Shock, n(%)	32 (18.82)	13 (9.92)	19 (48.72)	<.001
Respiratory failure, n(%)	164 (96.47)	127 (96.95)	37 (94.87)	0.903
ARDS, n(%)	12 (7.06)	9 (6.87)	3 (7.69)	1.000
Myocardial injury, n(%)	50 (29.41)	34 (25.95)	16 (41.03)	0.070
ECMO, n(%)	3 (1.76)	3 (2.29)	0 (0.00)	1.000
Hepatic injury, n(%)	34 (20.00)	29 (22.14) 5 (12.82)		0.202
AKI, n(%)	9 (5.29)	6 (4.58) 3 (7.69)		0.723
Other respiratory pathogen infections ^a , n(%)	129 (75.88)	100 (76.34)	29 (74.36)	0.800
Pneumonia, n(%)	165 (97.06)	128 (97.71)	37 (94.87)	0.703
Atelectasis, n(%)	66 (38.82)	54 (41.22)	12 (30.77)	0.240
Lung consolidation, n(%)	121 (71.18)	90 (68.70)	31 (79.49)	0.192
Vasoactive drugs, n(%)	98 (57.65)	69 (52.67)	29 (74.36)	0.016
Albumin, n(%)	124 (72.94)	86 (65.65)	38 (97.44)	<.001
Hormones, n(%)	98 (57.65)	78 (59.54)	20 (51.28)	0.359
IVIG, n(%)	110 (64.71)	86 (65.65) 24 (61.54)		0.637
Ventilator, n(%)	155 (91.18)	117 (89.31) 38 (97.44)		0.212
High frequency ventilation, n(%)	11 (6.47)	6 (4.58) 5 (12.82)		0.143
Invasive ventilation, n(%)	142 (83.53)	106 (80.92) 36 (92.31)		0.092
Mortality,n (%)	38 (22.35)	22 (16.79)	16 (41.03)	0.001

ALT Alanine transaminase, AKI Acute kidney injury, ARDS Acute respiratory distress syndrome, CRP C-reactive protein, ECMO Extracorporeal membrane oxygenation, IQR Interquartile range, IVIG Intravenous immune globulin, M Median, PCT Procalcitonin, PCO₂ Partial pressure of carbon dioxide, PO₂ Oxygen partial pressure, PLT Platelet, PH Pulmonary arterial hypertension, WBC White blood cells

^a Other respiratory pathogen infections include the following pathogens: Adenovirus, Epstein-Barr Virus, Bocavirus, Parainfluenza Virus, Coronavirus, Respiratory Syncytial Virus, Influenza Virus, Acinetobacter baumannii, Citrobacter freundii, Enterobacter cloacae, Escherichia coli, Klebsiella pneumoniae, Streptococcus pneumoniae, Haemophilus influenzae, Staphylococcus aureus, Moraxella catarrhalis, Enterobacter aerogenes, Serratia marcescens, Leminospira nodosa, Methicillin-Resistant Staphylococcus aureus, Elizabethkingia meningoseptica, Klebsiella oxytoca, Achromobacter xylosoxidans, Pseudomonas aeruginosa, Lysinibacillus sphaericus, Candida albicans, Chlamydia pneumoniae, Mycoplasma pneumoniae, Chlamydia trachomatis, and others

circulating PT levels and thereby mitigating other potentially fatal effects.

Since Romano et al. published the first report in 2004 on severe pertussis patients receiving double-volume

exchange transfusion, an increasing number of case reports and small-scale studies have focused on this therapeutic approach [25, 26]. Our data show that survivors undergoing exchange transfusion experienced a

Table 3	Characteristics o	f exchange transf	usion patients	according to outcome

Variables	Total (<i>n</i> = 39)	Survivors (n = 23)	Deceased ($n = 16$)	Р
Age (day), M (IQR)	71 (51, 192)	94(54, 244)	63 (40, 94)	0.050
Gender (male), n(%)	22 (56.41)	11 (47.83)	11 (68.75)	0.325
Immunization, n(%)	17 (43.59)	12 (52.17)	5 (31.25)	0.325
WBC (× 10 ⁹ /I),M (IQR)	66.12 (55.26, 83.10)	62.40 (50.43, 70.32)	80.06 (59.24, 92.89)	0.017
Lymphocytic absolute value, M (IQR)	28.22 (23.40, 37.78)	28.22 (22.88, 39.40)	27.41 (24.56, 34.91)	0.944
Neutrophils absolute value, M (IQR)	27.57 (20.33, 39.46)	22.47 (18.27, 31.67)	42.75 (28.19, 53.60)	<.001
Monocytes absolute value, M (IQR)	2.84 (2.12, 4.57)	2.45 (2.03, 3.98)	3.76 (2.27, 4.91)	0.199
PLT (× 10 ⁹ /l), M (IQR)	620 (527, 743)	620 (547, 743)	655(508, 742)	0.989
CRP (mg/l), M (IQR)	18 (8, 34)	8 (8, 22)	32 (16, 44)	0.019
PCT (ng/ml), M (IQR)	0.13 (0.08, 0.33)	0.10 (0.07, 0.26)	0.16 (0.12, 0.34)	0.106
Bilirubin (µmol/l), M (IQR)	6.30 (3.00, 13.15)	6.00 (1.95, 9.60)	9.45 (4.60, 18.77)	0.153
ALT (U/L), M (IQR)	30.60 (19.50, 42.70)	32.20 (21.00, 48.80)	24.50 (18.75, 33.50)	0.145
Creatinine (µmol/l), M (IQR)	21.00 (16.00, 25.20)	21.00 (16.00, 23.45)	19.55 (14.00, 26.50)	0.864
PCO ₂ (mmHg), M (IQR)	49.00 (40.40, 55.05)	43.40 (39.60, 53.50)	51.85 (42.70, 59.62)	0.242
PO2(mmHg), M (IQR)	78.20 (61.50, 97.20)	65.10 (58.00, 88.95)	87.05 (73.07, 163.25)	0.009
PH, n(%)	3 (7.69)	2 (8.70)	1 (6.25)	1.000
Epilepsy, n(%)	3 (7.69)	2 (8.70)	1 (6.25)	1.000
Hypoproteinemia, n(%)	16 (41.03)	8 (34.78)	8 (50.00)	0.509
Sepsis, n(%)	6 (15.38)	1 (4.35)	5 (31.25)	0.033
Shock, n(%)	19 (48.72)	4 (17.39)	15 (93.75)	<.001
Respiratory failure, n(%)	37 (94.87)	23 (100.00)	14 (87.50)	0.162
ARDS, n(%)	3 (7.69)	0 (0.00) 3 (18.75)		0.061
Myocardial injury, n(%)	16 (41.03)	8 (34.78) 8 (50.00)		0.509
Hepatic injury, n(%)	5 (12.82)	3 (13.04) 2 (12.50)		1.000
AKI, n(%)	3 (7.69)	0 (0.00)	3 (18.75)	0.061
Other respiratory pathogen infections ^a , n(%)	29 (74.36)	16 (69.57)	13 (81.25)	0.480
Atelectasis, n(%)	12 (30.77)	10 (43.48)	2 (12.50)	0.076
Lung consolidation, n(%)	31 (79.49)	19 (82.61)	12 (75.00)	0.694
Vasoactive drugs, n(%)	29 (74.36)	13 (56.52)	16 (100.00)	0.002
Albumin, n(%)	38 (97.44)	22 (95.65)	16 (100.00)	1.000
Hormones, n(%)	20 (51.28)	12 (52.17)	8 (50.00)	1.000
IVIG, n(%)	24 (61.54)	16 (69.57)	8 (50.00)	0.318
Ventilator, n(%)	38 (97.44)	22 (95.65) 16 (100.00)		1.000
High frequency ventilation, n(%)	5 (12.82)	2 (8.70)	3 (18.75)	0.631
Invasive ventilation, n(%)	36 (92.31)	21 (91.30)	15 (93.75)	1.000

ALT Alanine transaminase, AKI Acute kidney injury, ARDS Acute respiratory distress syndrome, CRP C-reactive protein, IQR Interquartile range, IVIG Intravenous immune globulin, M Median, PCT Procalcitonin, PCO₂ Partial pressure of carbon dioxide, PO₂ Oxygen partial pressure, PLT Platelet, PH Pulmonary arterial hypertension, WBC White blood cells

^a Other respiratory pathogen infections include the following pathogens: Adenovirus, Epstein-Barr Virus, Bocavirus, Parainfluenza Virus, Coronavirus, Respiratory Syncytial Virus, Influenza Virus, Acinetobacter baumannii, Citrobacter freundii, Enterobacter cloacae, Escherichia coli, Klebsiella pneumoniae, Streptococcus pneumoniae, Haemophilus influenzae, Staphylococcus aureus, Moraxella catarrhalis, Enterobacter aerogenes, Serratia marcescens, Leminospira nodosa, Methicillin-Resistant Staphylococcus aureus, Elizabethkingia meningoseptica, Klebsiella oxytoca, Achromobacter xylosoxidans, Pseudomonas aeruginosa, Lysinibacillus sphaericus, Candida albicans, Chlamydia pneumoniae, Mycoplasma pneumoniae, Chlamydia trachomatis, and others

dramatic reduction in white blood cell counts (from a median of $62.40 \times 10^{9}/L$ to $11.41 \times 10^{9}/L$), whereas the decrease was less pronounced in non-survivors (from $80.06 \times 10^{9}/L$ to $45.43 \times 10^{9}/L$). Furthermore, the death group exhibited persistently elevated

absolute neutrophil counts and CRP levels, suggesting that exchange transfusion may not fully address underlying risk factors such as secondary infections. These findings are consistent with previous reports [13, 20, 22]. A recent study reported that rapid proliferation

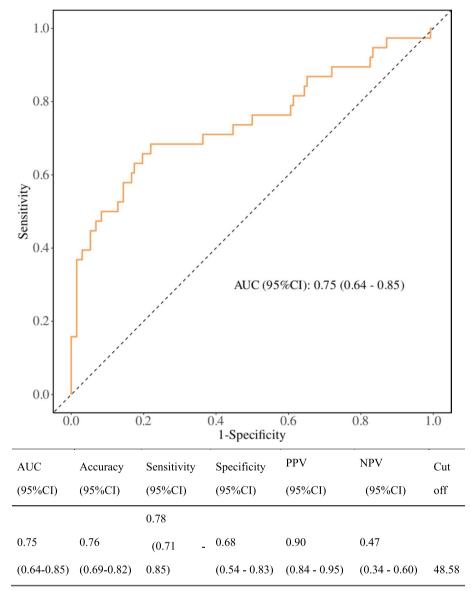


Fig. 1 Diagnostic Performance of WBC: ROC Curve Analysis for Mortality Prediction

Table 4	Univariate a	and multivariate	regression	analysis

Variables	Univariate			Multivariate		
	β	Р	OR (95%CI)	β	Р	OR (95%CI)
Immunization	0	0.997	1.00 (0.48 ~ 2.07)			
PH	- 0.91	0.24	0.40 (0.09 ~ 1.84)			
Sepsis	1.88	<.001	6.52 (2.70 ~ 15.73)	1.33	0.023	3.79 (1.21 ~ 11.91)
Shock	3.28	<.001	26.57 (10.05 ~ 70.26)	2.63	<.001	13.88 (4.72 ~ 40.82)
Myocardial injury	0.59	0.125	1.81 (0.85 ~ 3.85)			
Age (day)	0	0.315	1.00 (1.00 ~ 1.00)			
WBC ($\times 10^{9}$ /l)	0.04	<.001	1.04 (1.02 ~ 1.05)	0.02	0.029	1.02 (1.01 ~ 1.04)
PCO ₂ (mmHg)	0.03	0.049	1.03 (1.01 ~ 1.06)			

PCO₂ Partial pressure of carbon dioxide, WBC White blood cells, OR odds ratio, CI confidence interval

Variables	Survivors (n = 23)		Deceased (n = 16)	Deceased (n = 16)	
	Before treatment	last detection	Before treatment	last detection	
WBC (× 10 ⁹ /I),M (IQR)	62.40 (50.43, 70.32)	11.41 (10.05, 15.09)	80.06 (59.24, 92.89)	45.43 (26.12, 55.00)	
Lymphocytic absolute value, M (IQR)	28.22 (22.88, 39.40)	6.11 (5.20, 8.67)	27.41 (24.56, 34.91)	15.36 (5.41, 18.37)	
Monocytes absolute value, M (IQR)	2.45 (2.03, 3.98)	0.72 (0.56, 1.00)	3.76 (2.27, 4.91)	1.43 (0.82, 3.80)	
Neutrophils absolute value, M (IQR)	22.47 (18.27, 31.67)	3.85 (2.41, 5.09)	42.75 (28.19, 53.60)	23.82 (18.75, 32.74)	
PLT (× 10 ⁹ /l), M (IQR)	620 (547, 743)	591 (476, 715)	655 (508, 742)	153 (68, 249)	
CRP (mg/l), M (IQR)	8 (8, 22)	8(8, 8)	32 (16, 44)	37 (27, 68)	

Table 5 Characteristics of patients after exchange transfusion according to outcome

CRP C-reactive protein, IQR Interquartile range, M Median, PCT Procalcitonin, WBC White blood cells

of neutrophils during acute infection may exacerbate organ injury, which further explains the high neutro-phil levels observed in the death group [14].

Shock was significantly more prevalent in death group, consistent with reports linking pertussis-associated pulmonary hypertension to low cardiac output and fatal circulatory collapse [18, 24]. Other complications, including AKI and ARDS, were significantly more frequent in the death group, suggesting that pertussis-associated organ dysfunction may contribute to fatal outcomes [27]. Moreover, blood gas analysis revealed significant differences in PCO₂ between groups, indicating that CO₂ retention might further impact prognosis. Although the higher PO₂ observed in the death group likely reflects the early use of high-concentration oxygen therapy, this phenomenon merits additional investigation.

Finally, while the death group had a younger median age compared to survivors and a lower proportion received IVIG therapy, the influence of these factors on patient outcomes remains to be fully elucidated. Given the limited evidence regarding IVIG in pertussis management [28], further studies are necessary to determine its potential therapeutic role.

Limitations

Although this study provides valuable data on the clinical characteristics and mortality risk factors of children with severe pertussis, there are also several limitations. The study is retrospective, relying on clinical records that may contain omissions or inconsistencies, which could affect the accuracy of the results. In addition, as a single-center study, the external generalizability of the findings is limited. Even though the overall sample size is relatively large (170 cases), some subgroup analyses, such as the comparison between the exchange transfusion group and the non-exchange transfusion group, involve smaller samples that may reduce statistical significance and increase random error. Furthermore, the data were collected from January 2018 to June 2024, and changes in disease prevalence patterns during this period might have influenced the results.

Conclusion

Our study highlights the complex nature of severe pertussis, where hyperleukocytosis, secondary infections, and multi-organ complications collectively contribute to mortality. Notably, high WBC counts are significantly correlated with the risk of death, with ROC curve analysis indicating that when WBC counts exceed $48.58 \times 10^{\circ}/L$, the risk of mortality increases significantly. This suggests that WBC counts can serve as a valuable clinical indicator for identifying high-risk children and guiding early intervention. Secondary infections play a critical role in disease progression, emphasizing the need for strengthened infection control and timely intervention. While exchange transfusion may be beneficial for some severe children, its efficacy varies, as some patients remain at risk of death despite treatment. Therefore, its use should be carefully evaluated based on individual clinical conditions, particularly in children with high WBC counts. Future research should focus on establishing standardized thresholds for treatment initiation and developing integrated management strategies to improve patient outcomes.

Supplementary Information

The online version contains supplementary material available at https://doi. org/10.1186/s13052-025-01951-7.

Supplementary Material 1.

Acknowledgements None.

Authors' contributions

JH, CL and FX designed, supervised and revised the paper. JH also contributed to the data collection, data analysis, literature search, table, figures design and writing. SC contributed to the data analysis, literature search and table, figures

revisions. YQ participated in the database management and statistical analysis. All authors read and approved the final manuscript.

Funding

None.

Data availability

Data available on request from the authors. The data that support the findings of this study are available from the corresponding author, upon reasonable request.

Declarations

Ethics approval and consent to participate

This study was approved by the Ethics Committee (Institutional Review Board of Children's Hospital of Chongqing Medical University). The ethical considerations for this study are based on the guidelines provided by the World Medical Association (WMA) and the International Committee on Medical Ethics and Safety (CIOMS).

Consent for publication

N/A.

Competing interests

The authors have no relevant financial or non-financial interests to disclose.

Received: 4 February 2025 Accepted: 27 March 2025 Published online: 07 April 2025

References

- 1. Bamberger ES, Srugo I. What is new in pertussis? Eur J Pediatr. 2008;167:133–9. https://doi.org/10.1007/s00431-007-0548-2.
- Wood N, McIntyre P. Pertussis: review of epidemiology, diagnosis, management and prevention. Paediatr Respir Rev. 2008;9:201–12. https://doi. org/10.1016/j.prrv.2008.05.010.
- Yeung KHT, Duclos P, Nelson EAS, Hutubessy RCW. An update of the global burden of pertussis in children younger than 5 years: a modelling study. Lancet Infect Dis. 2017;17:974–80. https://doi.org/10.1016/S1473-3099(17)30390-0.
- Lashkari HP, Karuppaswamy S, Khalifa K. Pertussis-related hyperleukocytosis: role of hyperhydration and exchange transfusion. Clin Pediatr (Phila). 2012;51:987–90. https://doi.org/10.1177/0009922811410971.
- Straney L, Schibler A, Ganeshalingham A, Alexander J, Festa M, Slater A, et al. Burden and Outcomes of Severe Pertussis Infection in Critically III Infants. Pediatr Crit Care Med. 2016;17:735–42. https://doi.org/10.1097/ PCC.000000000000851.
- Winter K, Zipprich J, Harriman K, Murray EL, Gornbein J, Hammer SJ, et al. Risk Factors Associated With Infant Deaths From Pertussis: A Case-Control Study. Clin Infect Dis. 2015;61:1099–106. https://doi.org/10.1093/cid/ civ472.
- Mikelova LK, Halperin SA, Scheifele D, Smith B, Ford-Jones E, Vaudry W, et al. Predictors of death in infants hospitalized with pertussis: a case-control study of 16 pertussis deaths in Canada. J Pediatr. 2003;143:576–81. https://doi.org/10.1067/S0022-3476(03)00365-2.
- Kazantzi MS, Prezerakou A, Kalamitsou SN, Ilia S, Kalabalikis PK, Papadatos J, et al. Characteristics of Bordetella pertussis infection among infantsand children admitted to paediatric intensive care units in Greece: A multicentre, 11-year study. J Paediatr Child Health. 2017;53:257–62. https://doi. org/10.1111/jpc.13427.
- Kuperman A, Hoffmann Y, Glikman D, Dabbah H, Zonis Z. Severe pertussis and hyperleukocytosis: is it time to change for exchange? Transfusion. 2014;54:1630–3. https://doi.org/10.1111/trf.12519.
- Donoso AF, Cruces PI, Camacho JF, Leon JA, Kong JA. Exchange transfusion to reverse severe pertussis-induced cardiogenic shock. Pediatr Infect Dis J. 2006;25:846–8. https://doi.org/10.1097/01.inf.0000232630.70138.a2.
- 11. Martinez M, Rochat I, Corbelli R, Tissieres P, Rimensberger PC, Barazzone-Argiroffo C. Early blood exchange transfusion in malignant pertussis: a

case report. Pediatr Crit Care Med. 2011;12:e107–9. https://doi.org/10. 1097/PCC.0b013e3181f3a189.

- Rowlands HE, Goldman AP, Harrington K, Karimova A, Brierley J, Cross N, et al. Impact of rapid leukodepletion on the outcome of severe clinical pertussis in young infants. Pediatrics. 2010;126:e816–27. https://doi.org/ 10.1542/peds.2009-2860.
- Berger JT, Carcillo JA, Shanley TP, Wessel DL, Clark A, Holubkov R, et al. Critical pertussis illness in children: a multicenter prospective cohort study. Pediatr Crit Care Med. 2013;14:356–65. https://doi.org/10.1097/ PCC.0b013e31828a70fe.
- Coquaz-Garoudet M, Ploin D, Pouyau R, Hoffmann Y, Baleine JF, Boeuf B, et al. Malignant pertussis in infants: factors associated with mortality in a multicenter cohort study. Ann Intensive Care. 2021;11:70. https://doi.org/ 10.1186/s13613-021-00856-y.
- 15. Shi T, Wang L, Du S, Fan H, Yu M, Ding T, et al. Mortality risk factors among hospitalized children with severe pertussis. BMC Infect Dis. 2021;21:1057. https://doi.org/10.1186/s12879-021-06732-1.
- Namachivayam P, Shimizu K, Butt W. Pertussis: severe clinical presentation in pediatric intensive care and its relation to outcome. Pediatr Crit Care Med. 2007;8:207–11. https://doi.org/10.1097/01.PCC.0000265499.50592. 37.
- Mattoo S, Cherry JD. Molecular pathogenesis, epidemiology, and clinical manifestations of respiratory infections due to Bordetella pertussis and other Bordetella subspecies. Clin Microbiol Rev. 2005;18:326–82. https:// doi.org/10.1128/CMR.18.2.326-382.2005.
- Wang C, Zhang H, Zhang Y, Xu L, Miao M, Yang H, et al. Analysis of clinical characteristics of severe pertussis in infants and children: a retrospective study. BMC Pediatr. 2021;21:65. https://doi.org/10.1186/ s12887-021-02507-4.
- Liu C, Yang L, Cheng Y, Xu H, Xu F. Risk factors associated with death in infants <120 days old with severe pertussis: a case-control study. BMC Infect Dis. 2020;20:852. https://doi.org/10.1186/s12879-020-05535-0.
- Hodge G, Hodge S, Markus C, Lawrence A, Han P. A marked decrease in L-selectin expression by leucocytes in infants with Bordetella pertussis infection: leucocytosis explained? Respirology. 2003;8:157–62. https://doi. org/10.1046/j.1440-1843.2003.00459.x.
- Carbonetti NH. Pertussis leukocytosis: mechanisms, clinical relevance and treatment. Pathog Dis, (2016) 74:https://doi.org/10.1093/femspd/ftw087.
- Sawal M, Cohen M, Irazuzta JE, Kumar R, Kirton C, Brundler MA, et al. Fulminant pertussis: a multi-center study with new insights into the clinicopathological mechanisms. Pediatr Pulmonol. 2009;44:970–80. https://doi. org/10.1002/ppul.21082.
- Paddock CD, Sanden GN, Cherry JD, Gal AA, Langston C, Tatti KM, et al. Pathology and pathogenesis of fatal Bordetella pertussis infection in infants. Clin Infect Dis. 2008;47:328–38. https://doi.org/10.1086/589753.
- Goulin GD, Kaya KM, Bradley JS. Severe pulmonary hypertension associated with shock and death in infants infected with Bordetella pertussis. Crit Care Med. 1993;21:1791–4. https://doi.org/10.1097/00003246-19931 1000-00033.
- Romano MJ, Weber MD, Weisse ME, Siu BL. Pertussis pneumonia, hypoxemia, hyperleukocytosis, and pulmonary hypertension: improvement in oxygenation after a double volume exchange transfusion. Pediatrics. 2004;114:e264–6. https://doi.org/10.1542/peds.114.2.e264.
- Onoro G, Salido AG, Martinez IM, Cabeza B, Gillen M, de Azagra AM. Leukoreduction in patients with severe pertussis with hyperleukocytosis. Pediatr Infect Dis J. 2012;31:873–6. https://doi.org/10.1097/INF.0b013 e31825ba6cf.
- Nicholson CE. Critical pertussis may model organ failure in critical illness and injury. Pediatr Crit Care Med. 2007;8:288–9. https://doi.org/10.1097/ 01.PCC.0000265500.59962.B8.
- Pierce C, Klein N, Peters M. Is leukocytosis a predictor of mortality in severe pertussis infection? Intensive Care Med. 2000;26:1512–4. https:// doi.org/10.1007/s001340000587.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.